

# **Liquid Chromatography**

- 1. Introduction and Column Packing Material**
- 2. Retention Mechanisms in Liquid Chromatography**
- 3. Method Development**
- 4. Column Preparation**
- 5. General Instrumental aspects**
- 6. Detectors**

**(Chapter 4 and 5 in The essence of chromatography)**

# **Column Preparation**

**1. Column is the central part of liquid chromatography.**

## **2. Column Packing Methods**

**A. Dry-Packing Procedures: for rigid particles with diameter greater than 20  $\mu\text{m}$ .**

**B. Down-Fill Slurry Packing: for rigid particles with diameter smaller than 20  $\mu\text{m}$ .**

**C. Up-Fill Slurry Packing: for rigid particles with diameter smaller than 20  $\mu\text{m}$ .**

## Down-Fill Slurry Packing

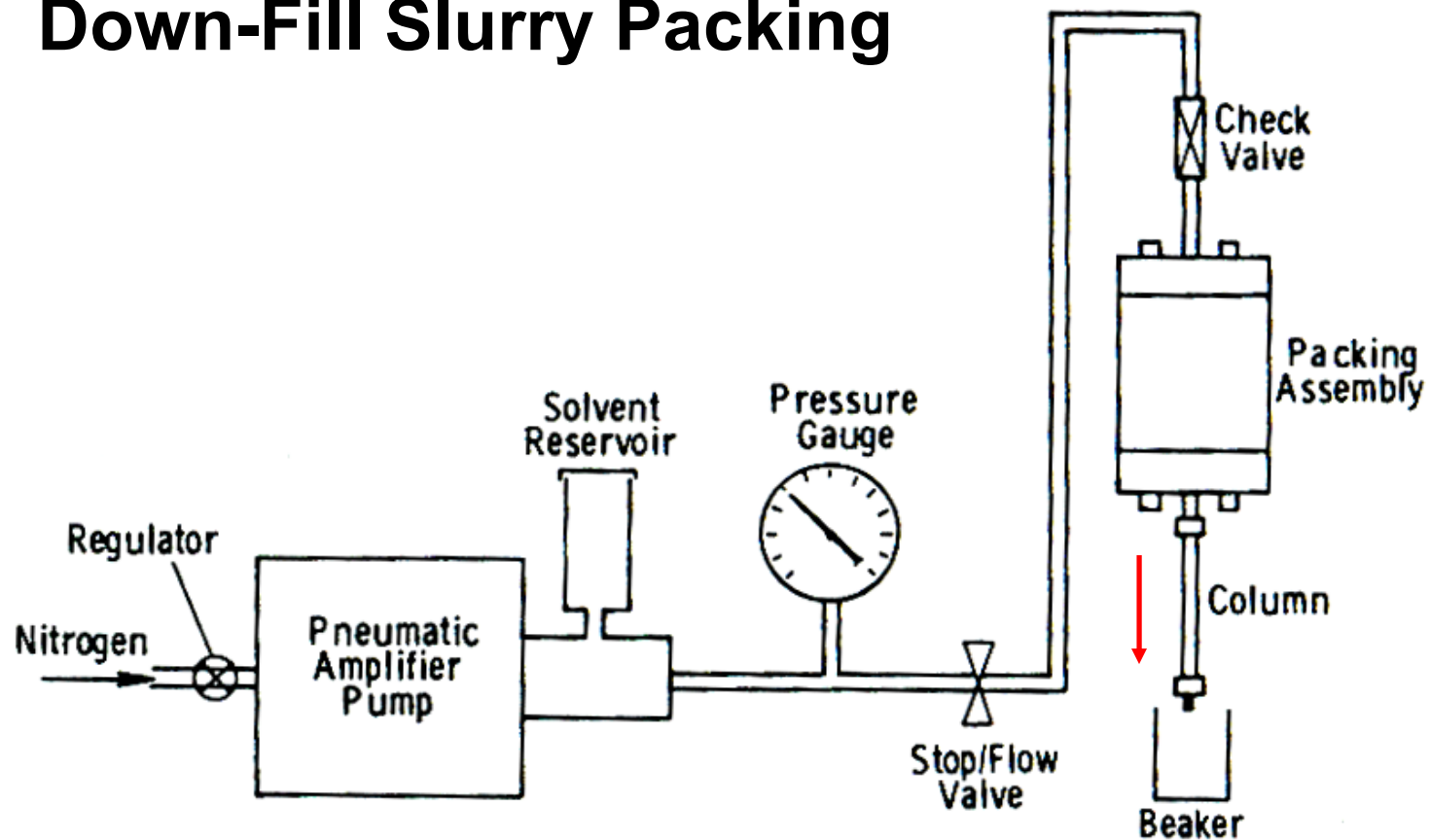
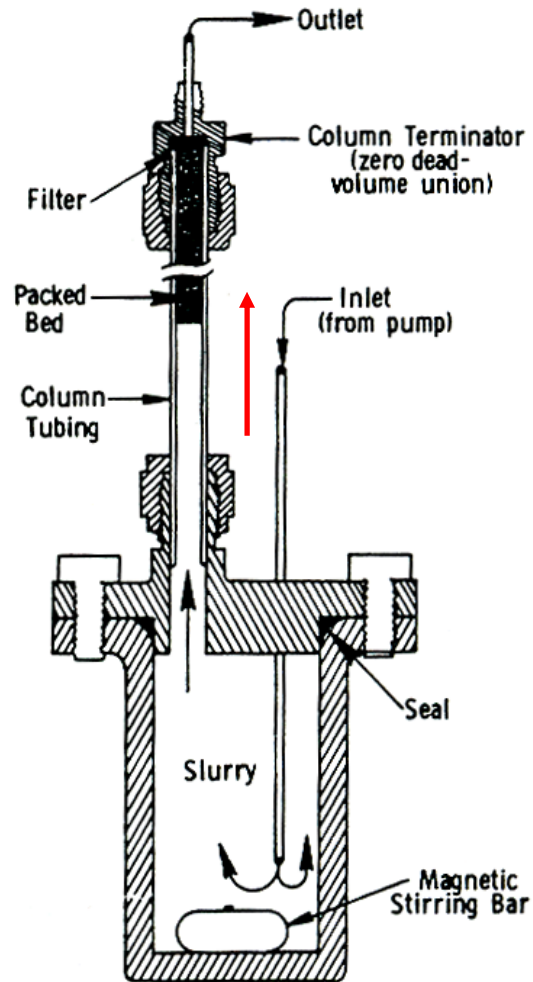


Figure 4.31. Down-fill slurry packing apparatus. (From ref. [658]. ©Elsevier)

# Up-Fill Slurry Packing



**This method is used for conventional Diameter columns. It is unsuitable for packing small diameter columns.**

Figure 4.32. Up-fill slurry packing apparatus.

### 3. Evaluation of column quality

Table 4.19

Test mixtures for routine quality evaluation of normal- and reversed-phase columns

---

<i>Normal-phase columns</i>	
(1)	Mixture: toluene, nitrobenzene and p-nitroaniline Mobile phase: isooctane-ethanol-water (84.5:15:0.5 v/v)
(2)	Mixture: naphthalene, m-dinitrobenzene and o-nitroaniline Mobile phase: hexane-dichloromethane-2-propanol (89.5:10:0.5 v/v)
(3)	Mixture: toluene, phenanthrene and nitrobenzene Mobile phase: hexane-acetonitrile (99:1 v/v)
(4)	Mixture: toluene, nitrobenzene, acetophenone, 2,6-dinitrotoluene and 1,3,5-trinitrobenzene Mobile phase: hexane-methanol (99.5:0.5 v/v)
<i>Reversed-phase columns</i>	
(1)	Mixture: resorcinol, acetophenone, naphthalene and anthracene Mobile phase: acetonitrile-water (55:45 v/v)
(2)	Mixture: uracil, phenol, benzaldehyde, N,N-dimethyl-3-toluamide, toluene and ethylbenzene Mobile phase: acetonitrile-water (65:35 v/v)
(3)	Mixture: acetone, acetophenone, anisole, benzene and toluene Mobile phase: acetonitrile-water (60:40 v/v)
(4)	Mixture: thiourea, phenol, 1-chloro-4-nitrobenzene, toluene, ethylbenzene, n-butylbenzene Mobile phase: methanol-water (80:20 v/v)
(5)	Mixture: uracil, toluene, acenaphthene, propylparaben, dipropylphthalate Mobile phase: methanol-water (65:35 v/v)

---

**Major parameters: Capacity factor, plate numbers, separation factor, and asymmetry factor.**

# Routine Column Quality Evaluation

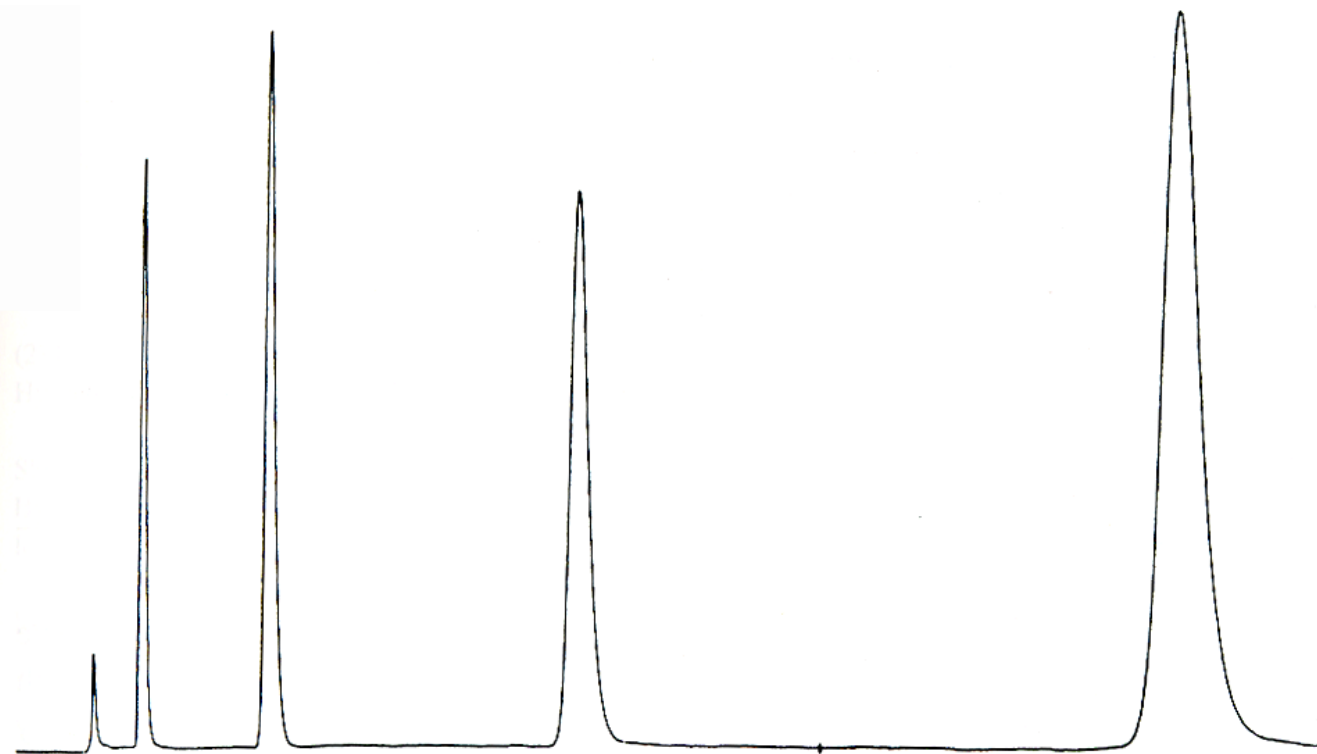
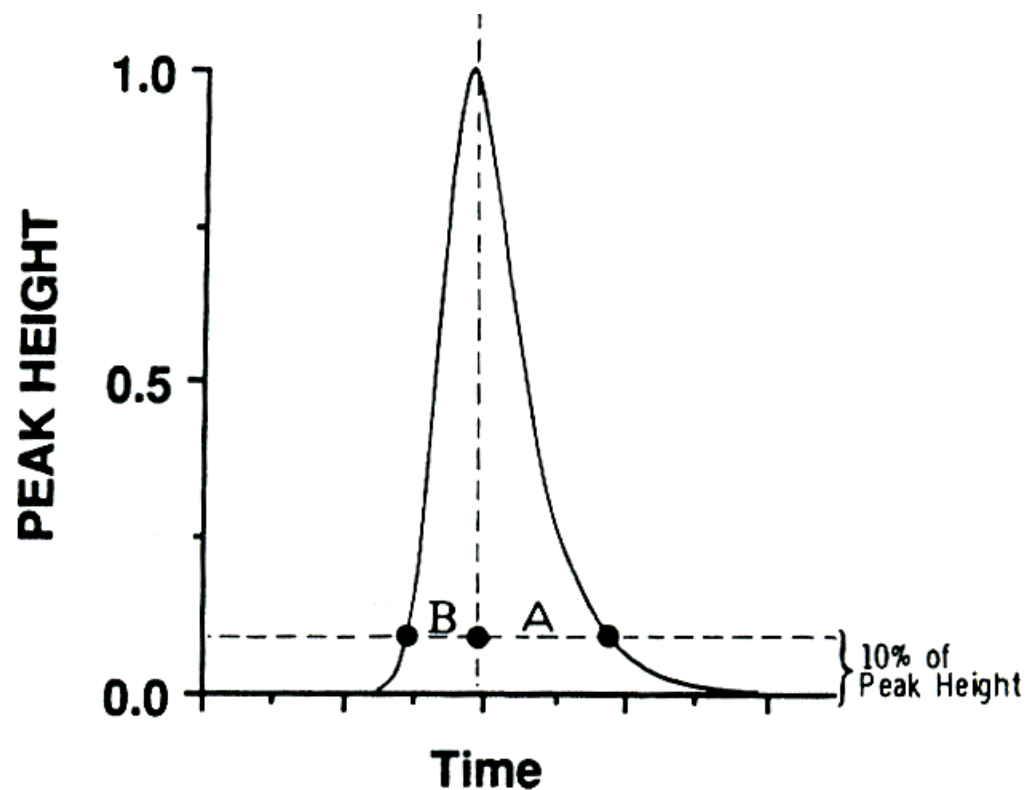


Figure 4.33. Typical routine column quality test chromatogram for a 30 cm x 4.6 mm column packed with ar octadecylsiloxane-bonded silica packing of 10  $\mu$ m particle diameter. Test mixture: resorcinol (0.55 mg / ml) acetophenone (0.025 mg / ml), naphthalene (0.20 mg/ml) and anthracene (0.01 mg / ml) in acetonitrile, 10  $\mu$  injected. Isocratic separation at 23°C with acetonitrile-water (55:45) as the mobile phase with a flow rate o 1.5 ml / min.

Solute	Retention factor	Asymmetry factor	N / m	Separation factor
Resorcinol	0.2	1.00	10,725	
Acetophenone	1.4	1.08	15,900	
Naphthalene	4.3	1.20	17,875	3.30
Anthracene	9.8	1.40	18,380	2.34

# Asymmetry Factor



$$\text{Asymmetry factor} = \frac{A}{B}$$

# Specific column quality test for chemically bonded phases

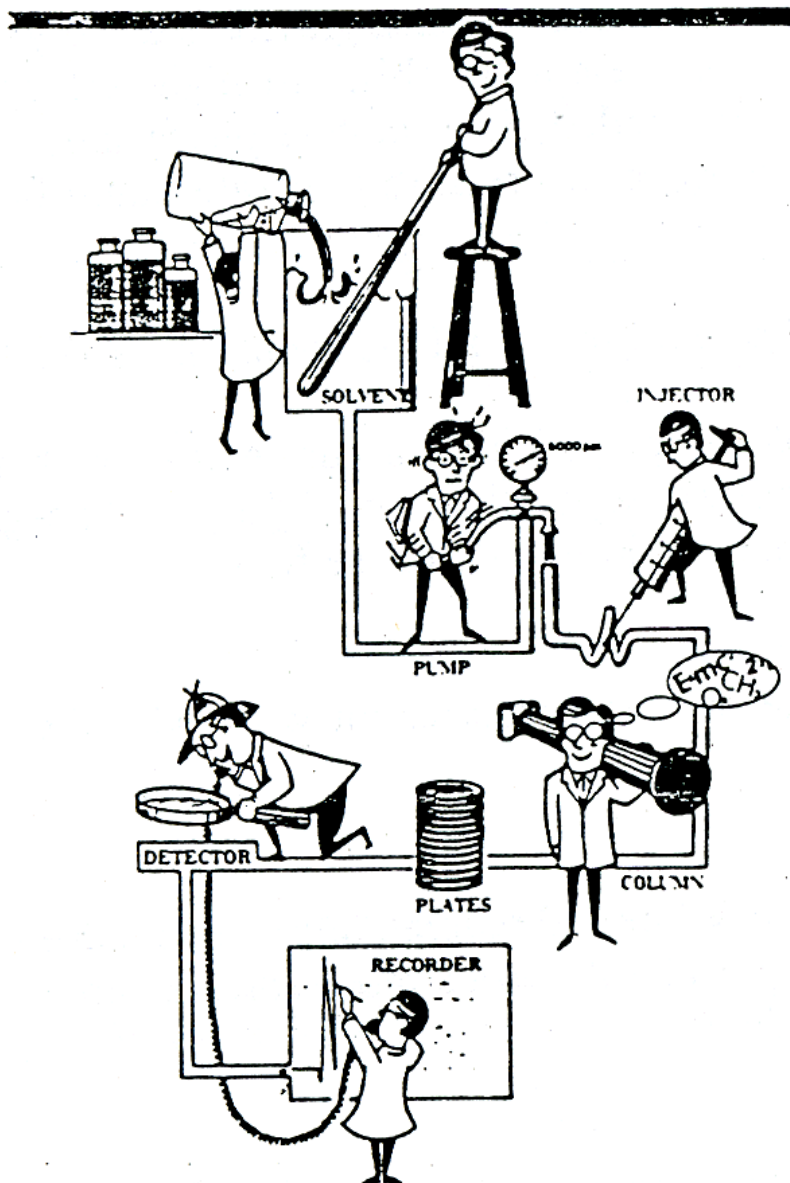
Table 4.21

Mixtures for specific property tests of reversed-phase columns

$\alpha$  = separation factor,  $k$  = retention factor and  $A_s$  = asymmetry factor

Property	Measurement
(1) Walters [660]	
Hydrophobicity	$\alpha$ (anthracene / benzene) with acetonitrile-water (65:35)
Silanophilicity	$\alpha$ (N,N-diethyltoluamide / anthracene) with acetonitrile $k$ (nitrobenzene) with n-heptane
(2) Kimata et al [662]	
Hydrophobicity	$k$ for n-pentylbenzene with methanol-water (80:20) $\alpha$ (n-pentylbenzene / n-butylbenzene) with methanol-water (80:20)
Shape selectivity	$\alpha$ (triphenylene / o-terphenyl) with methanol-water (80:20)
Hydrogen-bonding	$\alpha$ (caffeine / phenol) with methanol-water (30:70)
Ion exchange (pH >7)	$\alpha$ (benzylamine / phenol) with methanol-0.02M phosphate buffer pH 7.6 (30:70)
Ion exchange (pH <3)	$\alpha$ (benzylamine / phenol) with methanol-0.02M phosphate buffer pH 2.7 (30:70)
(3) Sander and Wise [75,76,669]	
Shape selectivity	$\alpha$ (1,2:3,4:5,6:7,8-tetrabenzonaphthalene / benzo[a]pyrene) with acetonitrile-water (85:15)
(4) Engelhardt and Lobert [670]	
Metal impurities	$100A_s(2,2'$ -bipyridyl) / $A_s(4,4'$ -bipyridyl) with methanol-water (49:51)
(5) Cruz et al [664]	
Metal impurities	Base peak efficiency (2,7-dihydroxynaphthalene) / base peak efficiency (2,3-dihydroxynaphthalene) with acetonitrile-25 mM ammonium acetate buffer pH 7.2 (25:75). 2,3-Dihydroxynaphthalene (300 mg / l) and 2,7-dihydroxynaphthalene (150 mg / l).
<i>Comprehensive test mixtures</i>	
Neue and co-workers [665]	
Uracyl (16 mg / l), toluene (300 $\mu$ l / l) or naphthalene (60 mg / l), acenaphthene (200 mg / l), propylparaben or butylparaben (20 mg / l), dipropylphthalate (300 mg / l) or dibutylphthalate (400 mg / l), propranolol (400 mg / l) and amitriptyline (100 mg / l) or doxepin (100 mg / l) with methanol-20 mM phosphate buffer pH 7 (65:35 v/v).	
Engelhardt and co-workers [671,672]	
Thiourea (12 mg / l), toluene (870 mg / l), ethylbenzene (867 mg / l), ethylbenzoate (523 mg / l), aniline (81.7 mg / l), o-touidine (79.8 mg / l), p-toluidine (20 mg / l), N,N-dimethylaniline (38.2 mg / l), phenol (120 mg / l) with methanol-water (55:45)	

# General Instrumental aspects

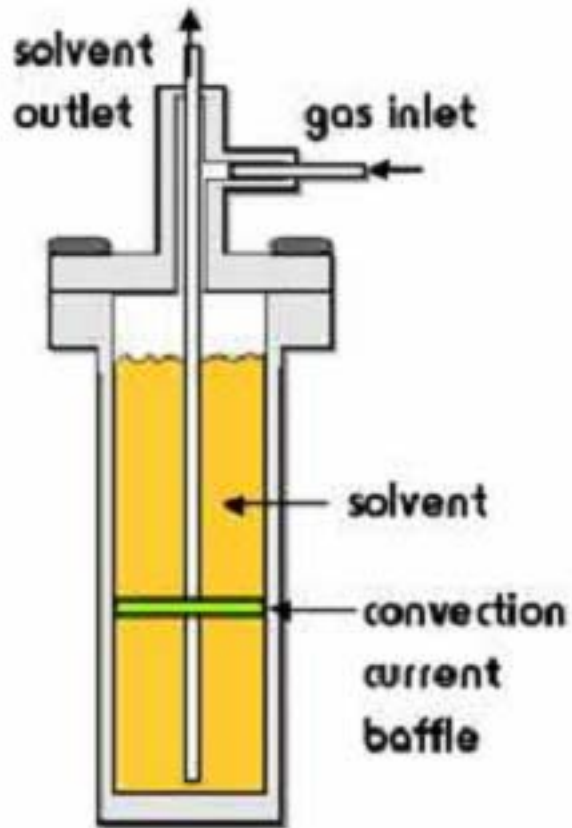


1. Pumps

2. Injectors

3. Detectors

# Direct pressure pump



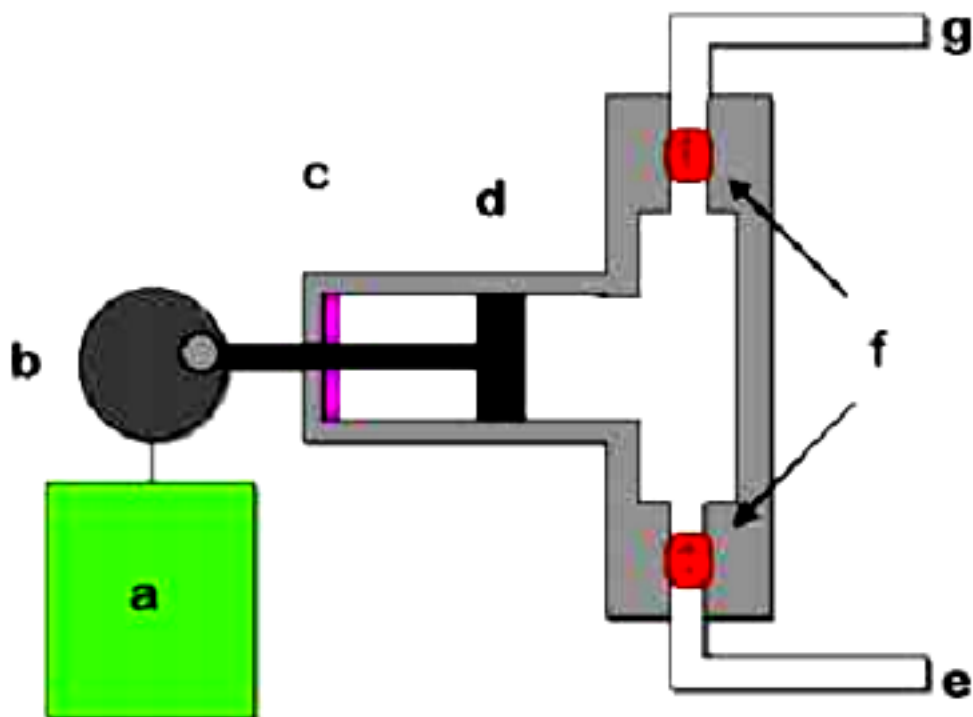
Gas pressure is applied from an external gas tank using a high Pressure regulator.

No pressure pulses are produced

The solvent reservoir is limited

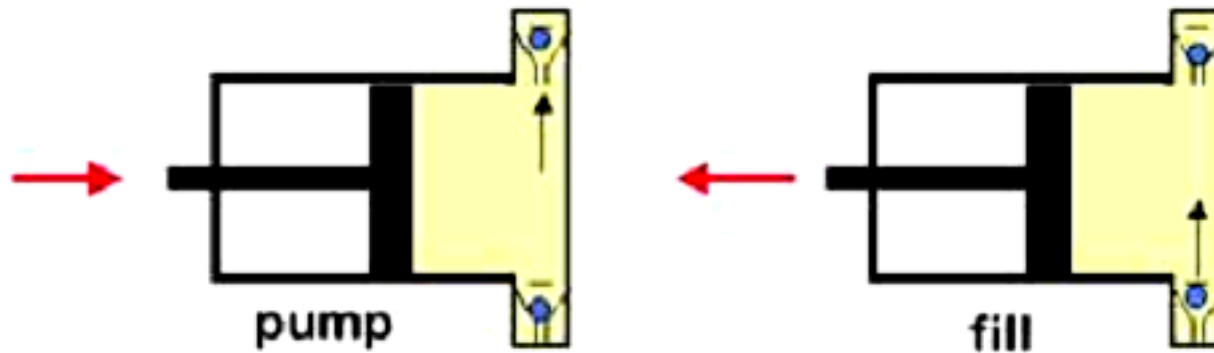
A major problem is introduction Of gas into the solvent.

## Reciprocating Pump



- a. Motor
- b. Gear
- c. Seal
- d. Piston
- e. Solvent in
- f. Check valves
- g. Solvent out

# Reciprocating Pump

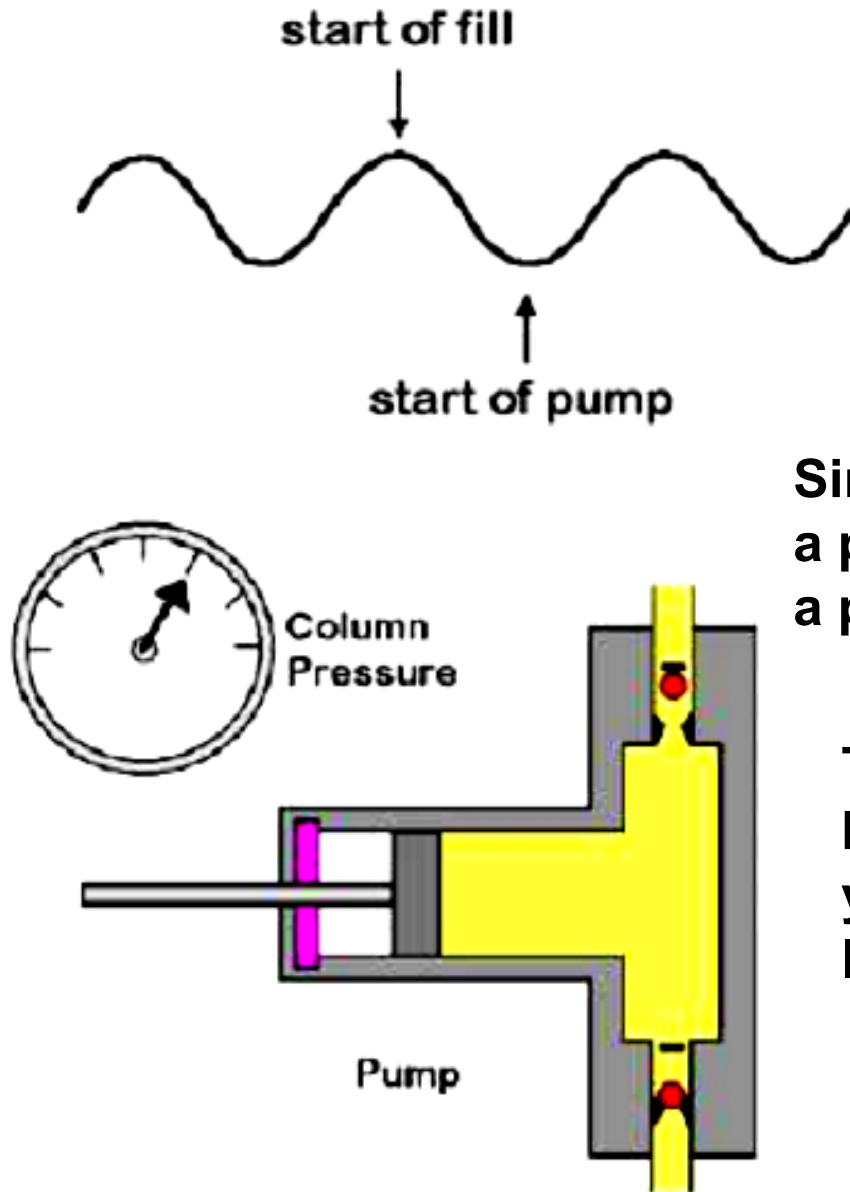


One of the most common type of system

Unlimited reservoir system

A major problem is that it produces variable pressure

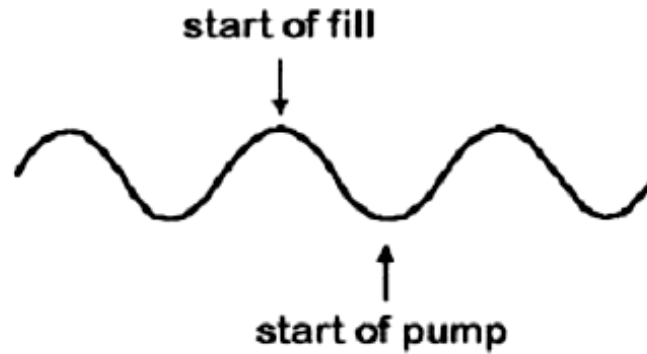
# Pressure Variances



Since the pump must spend at least a portion of its time filling, there is a pressure drop during the phase.

This effect must be minimized because it would greatly affect your sensitivity and detection limit.

# Reciprocating Pump



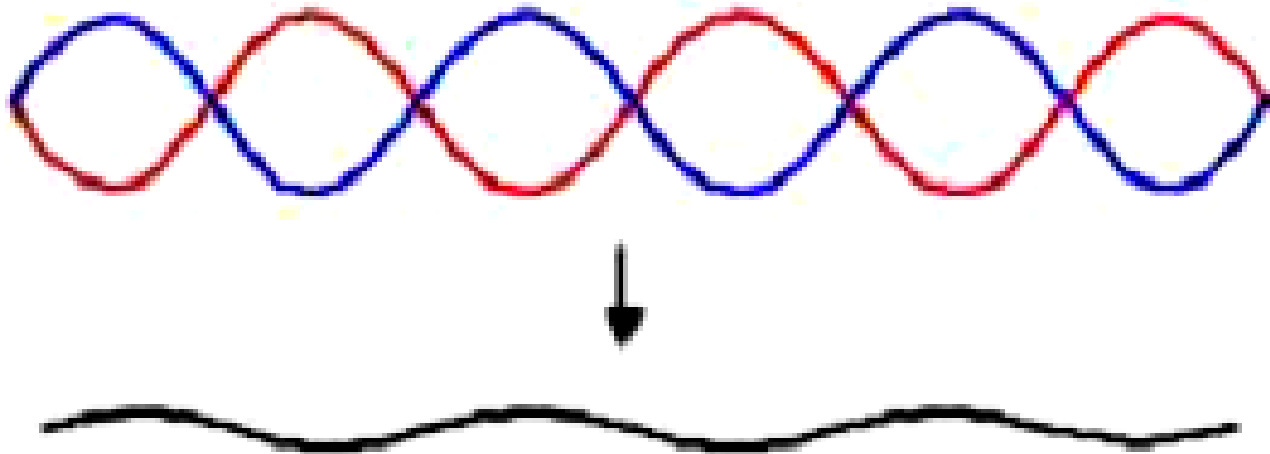
**One approach is to have a more rapid fill cycle**



**This does not eliminate the problem by reduce it**

# Reciprocating Pump

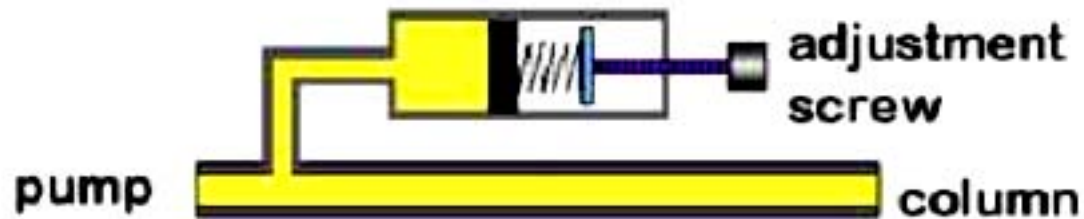
One could also use two or more pumps working in tandem



# Pulse Dampers

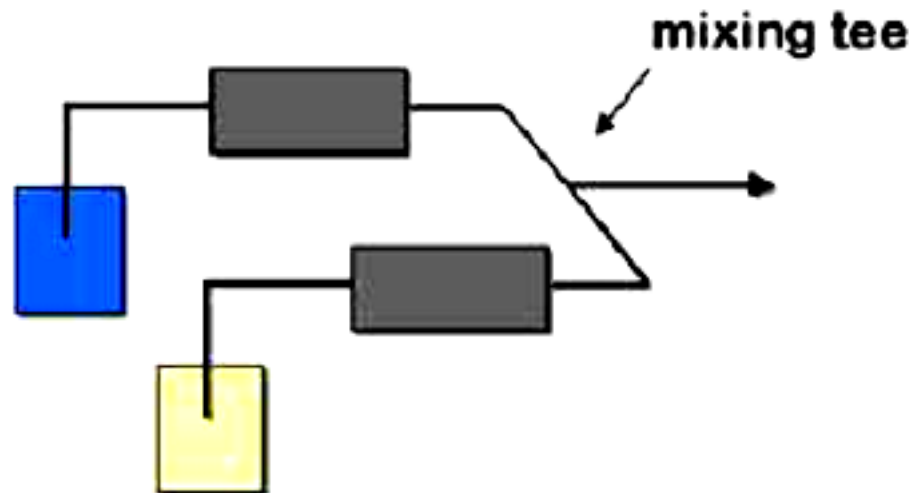
**The Popular Approach to minimize flow pulse associated with Reciprocating pumps**

**Mechanism: Absorb the peak and valley of the pressure pulse**



# Gradient Controller

## Dual pumping systems



A valve system can be used on each pump that can provide a different solvent.

# Valve Injectors

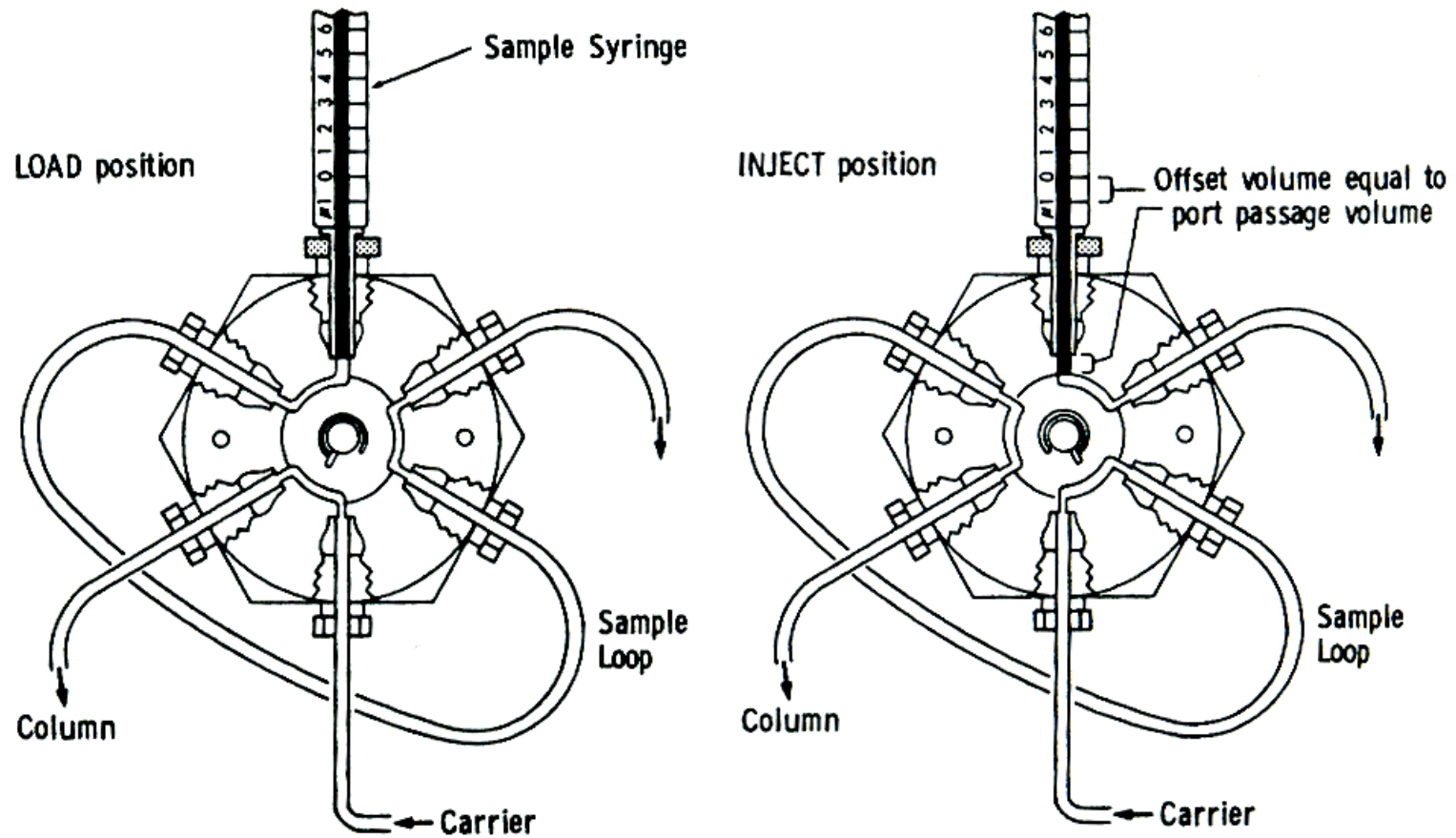
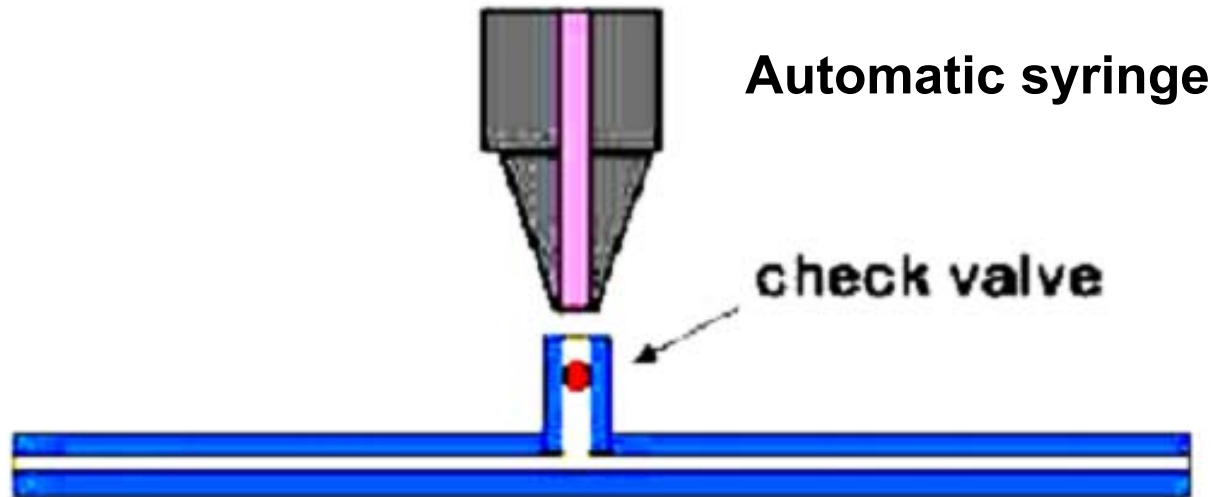


Figure 5.5. Sample injection valve showing the valve configuration in the load and inject position. (Reproduced with permission from Valco Instruments, Inc.).

# Automatic Injectors



**This method allow for adjustment of sample size.  
The motor driven syringe can provide sufficient pressure  
To inject sample past the check valve.**

# Trace Enrichment Sampling

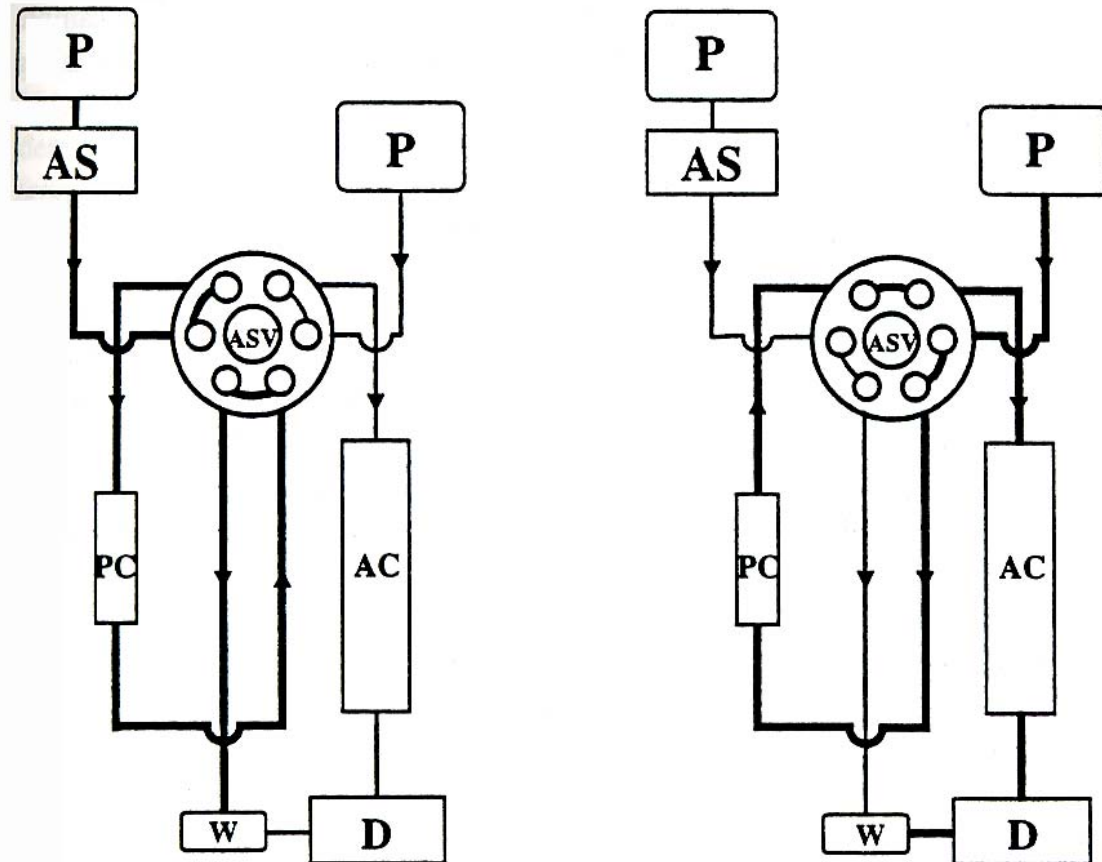


Figure 5.6. Arrangement for automated coupled-column switching in liquid chromatography. Position (I) is for sample application and fractionation and position (II) for transfer of extracted analytes and subsequent separation. P = pump, AS = autosampler, ASV = automated switching valve, PC = precolumn, AC = analytical column, D = detector and W = waste. (From ref. [41]; ©Elsevier).

# Multi-Dimensional LC

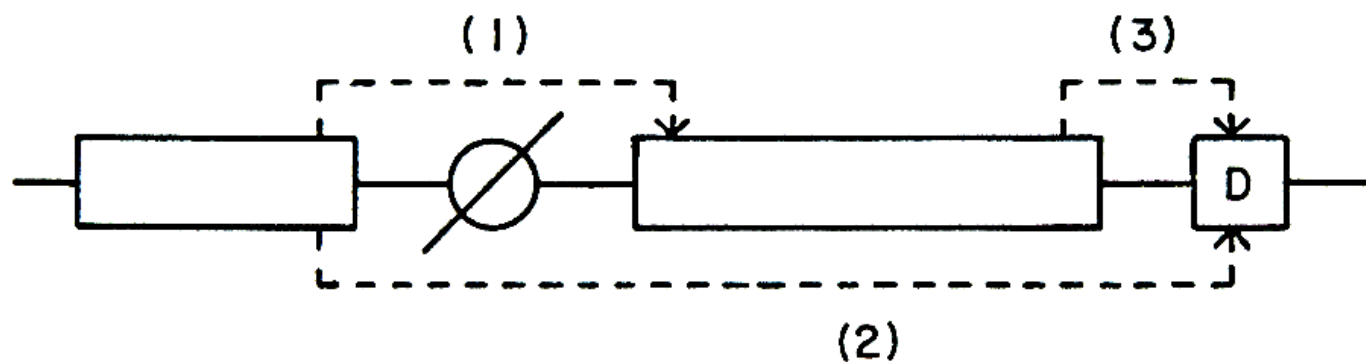


Figure 5.8. Schematic diagram of a two-column system for the separation of a sample containing components with a wide range of retention factors.

**Dimension -- column**

## 2-D Liquid Chromatography

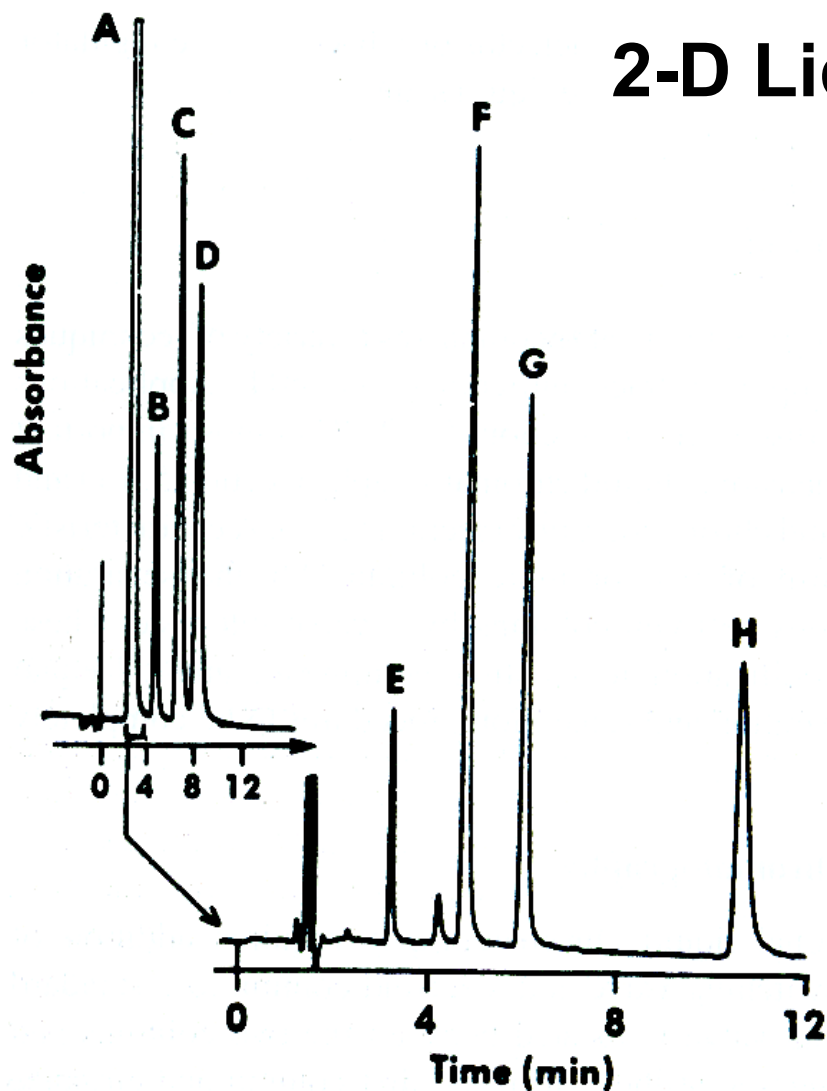
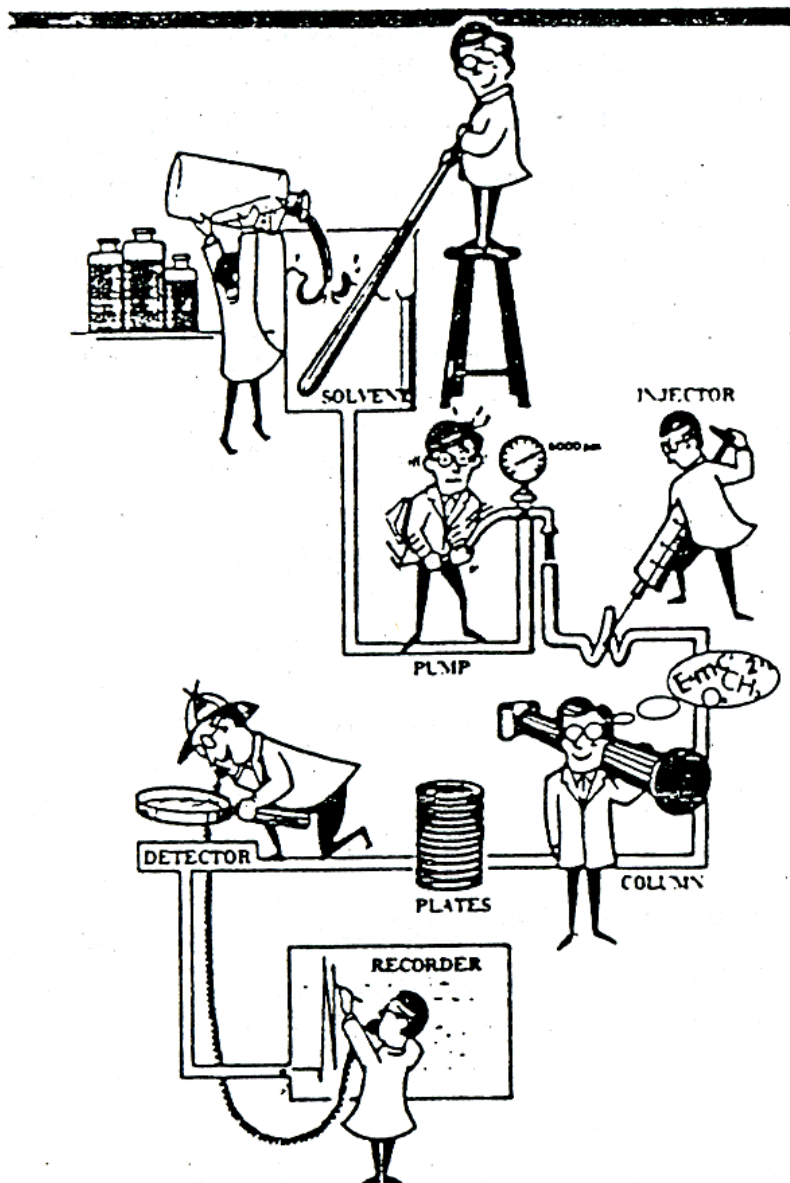


Figure 5.7. Separation of deoxyribonucleosides and their 5'- monophosphate esters by multidimensional liquid chromatography on a strong cation-exchange column (column one) and a reversed-phase column. The unseparated nucleosides, peak A, on the ion-exchange column were switched to the reversed-phase column for separation. Peak identification: A = nucleosides, B = d-CMP, C = d-AMP, D = d-GMP, E = d-CYD, F = d-URD, G = THD, and H = d-ADO. (From ref. [84]; ©Preston Publications, Inc.).

# General Instrumental aspects



1. Pumps
2. Injectors
3. Detectors

**1. Solute 1 and 2 are eluted from a reversed-phase column with retention time of 18.1 min and 23.5 min at a flow-rate of 1 mL/min, using a 10% 2-propanol: 90%water mixture as the mobile phase. The void time of the column at this flow-rate is 0.9 min. The strength of 2-propanol and water are 3.9 and 0, respectively.**

- a. What % 2-propanol must be used in order to elute 1 with a retention time of 10.0 min? What will be the retention time of solute 2 under this condition.**
- b. What mixture of water and THF ( $S' = 4.4$ ) will allow elution of 1 with a retention time of 10.0 mins?**